



## The Role of Antibiotic-Target-Modifying and Antibiotic-Modifying Enzymes in Drug Resistance

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**Abstract:** The incidence and prevalence of non-tuberculous mycobacterial (NTM) infections have been increasing worldwide and lately led to an emerging public health problem. Among rapidly growing NTM, is the most pathogenic and drug resistant opportunistic germ, responsible for disease manifestations ranging from "curable" skin infections to only "manageable" pulmonary disease. Challenges in treatment stem from the bacteria's high-level innate resistance and comprise long, costly and non-standardized administration of antimicrobial agents, poor treatment outcomes often related to adverse effects and drug toxicities, and high relapse rates. Drug resistance in is conferred by an assortment of mechanisms. Clinically acquired drug resistance is normally conferred by mutations in the target genes. Intrinsic resistance is attributed to low permeability of cell envelope as well as to (multi)drug export systems. However, expression of numerous enzymes by , which can modify either the drug-target or the drug itself, is the key factor for the pathogen's phenomenal resistance to most classes of antibiotics used for treatment of other moderate to severe infectious diseases, like macrolides, aminoglycosides, rifamycins, -lactams and tetracyclines. In 2009, when genome sequence became available, several research groups worldwide started studying antibiotic resistance mechanisms. At first, lack of tools for genetic manipulation severely delayed research endeavors. Nevertheless, the last 5 years, significant progress has been made towards the development of conditional expression and homologous recombination systems for . As a result of recent research efforts, an erythromycin ribosome methyltransferase, two aminoglycoside acetyltransferases, an aminoglycoside phosphotransferase, a rifamycin ADP-ribosyltransferase, a -lactamase and a monooxygenase were identified to frame the complex and multifaceted intrinsic resistome of , which clearly contributes to complications in treatment of this highly resistant pathogen. Better knowledge of the underlying mechanisms of drug resistance in could improve selection of more effective chemotherapeutic regimen and promote development of novel antimicrobials which can overwhelm the existing resistance mechanisms. This article reviews the currently elucidated molecular mechanisms of antibiotic resistance in , with a focus on its drug-target-modifying and drug-modifying enzymes.

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# The Role of Antibiotic-Target-Modifying and Antibiotic-Modifying Enzymes in *Mycobacterium abscessus* Drug Resistance

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The incidence and prevalence of non-tuberculous mycobacterial (NTM) infections have been increasing worldwide and lately led to an emerging public health problem. Among rapidly growing NTM, *Mycobacterium abscessus* is the most pathogenic and drug resistant opportunistic germ, responsible for disease manifestations ranging from “curable” skin infections to only “manageable” pulmonary disease. Challenges in *M. abscessus* treatment stem from the bacteria’s high-level innate resistance and comprise long, costly and non-standardized administration of antimicrobial agents, poor treatment outcomes often related to adverse effects and drug toxicities, and high relapse rates. Drug resistance in *M. abscessus* is conferred by an assortment of mechanisms. Clinically acquired drug resistance is normally conferred by mutations in the target genes. Intrinsic resistance is attributed to low permeability of *M. abscessus* cell envelope as well as to (multi)drug export systems. However, expression of numerous enzymes by *M. abscessus*, which can modify either the drug-target or the drug itself, is the key factor for the pathogen’s phenomenal resistance to most classes of antibiotics used for treatment of other moderate to severe infectious diseases, like macrolides, aminoglycosides, rifamycins,  $\beta$ -lactams and tetracyclines. In 2009, when *M. abscessus* genome sequence became available, several research groups worldwide started studying *M. abscessus* antibiotic resistance mechanisms. At first, lack of tools for *M. abscessus* genetic manipulation severely delayed research endeavors. Nevertheless, the last 5 years, significant progress has been made towards the development of conditional expression and homologous recombination systems for *M. abscessus*. As a result of recent research efforts, an erythromycin ribosome methyltransferase, two aminoglycoside acetyltransferases, an aminoglycoside phosphotransferase, a rifamycin ADP-ribosyltransferase, a  $\beta$ -lactamase and a monooxygenase were identified to frame the complex and multifaceted intrinsic resistome of *M. abscessus*, which clearly contributes to complications in treatment of this highly resistant pathogen. Better knowledge of the underlying mechanisms of drug resistance in *M. abscessus* could improve selection of more effective chemotherapeutic regimen and promote

development of novel antimicrobials which can overwhelm the existing resistance mechanisms. This article reviews the currently elucidated molecular mechanisms of antibiotic resistance in *M. abscessus*, with a focus on its drug-target-modifying and drug-modifying enzymes.

**Keywords:** non-tuberculous mycobacteria, *Mycobacterium abscessus*, antibiotic, drug resistance, resistance genes, antibiotic-target-modifying enzymes, antibiotic-modifying enzymes

## INTRODUCTION

Non-tuberculous mycobacteria (NTM) encompass all species of mycobacteria that do not cause tuberculosis (TB) or leprosy (Lee et al., 2015). NTM are ubiquitous in the environment and occasionally infect humans with various predisposing conditions like cystic fibrosis, bronchiectasis or immunosuppression, causing a variety of pathological conditions including pulmonary, skin and soft tissue infections and disseminated diseases (Chan and Iseman, 2013). The last few decades saw an alarming increase in the incidence and prevalence of NTM-pulmonary disease throughout the globe. Importantly, more than 90% of all reported NTM-pulmonary disease cases involved infections with the *Mycobacterium avium* complex (comprising of *M. avium*, *Mycobacterium chimaera* and *Mycobacterium intracellulare*) or *Mycobacterium abscessus*, accentuating the medical importance of these emerging pathogens (Hoefsloot et al., 2013; Ryu et al., 2016; Wu et al., 2018). For a long while, it was believed that infections with genetically diverse NTM strains were exclusively acquired upon exposure to the environment. However, a recent whole genome analysis of more than 1000 *M. abscessus* clinical isolates from different geographical locations, revealed the presence of genetically clustered strains in patients. This suggests a human-to-human transmission of *M. abscessus*, presumably through indirect mechanisms like cough aerosols or surface contamination (Bryant et al., 2016).

*Mycobacterium abscessus* is an opportunistic pathogen, ubiquitous in the environment, that often causes infections in humans with compromised natural defenses such as patients with cystic fibrosis or other chronic lung diseases (Sanguinetti et al., 2001; Brown-Elliott and Wallace, 2002; Howard, 2006; Griffith et al., 2007). A current taxonomic classification suggests separation of *M. abscessus* into three distinct subspecies: *M. abscessus* subsp. *abscessus*, *M. abscessus* subsp. *bolletii*, and *M. abscessus* subsp. *massiliense* (Tortoli et al., 2016). Although a saprophyte in water and soil, following lung infection *M. abscessus* can swiftly grow and survive intra-cellularly within macrophages as well as in extra-cellular caseous lesions and airway mucus (Wu et al., 2018). Several factors contribute to the success of this rapidly growing mycobacterium. A plethora of intrinsic resistance mechanisms renders almost all clinically used antibiotics ineffective against *M. abscessus* (Nessar et al., 2012). In addition, the presence of a highly dynamic open pan-genome in *M. abscessus* might explain the ease with which the bacterium evolves and adapts to a wide-spectrum of stressful environmental conditions encountered in diverse habitats (Choo et al., 2014; Wu et al., 2018). Importantly, the respiratory habitat of *M. abscessus* brings it in close proximity to highly virulent pathogens (for

example, *Pseudomonas aeruginosa* in cystic fibrosis lung) which can serve as donors of novel drug resistance or virulence genes (Ripoll et al., 2009).

Treatment of an *M. abscessus* pulmonary infection is very difficult owing to the bacterium's high-level innate resistance towards most antibiotics commonly used for Gram-negative and Gram-positive bacterial infections including majority of  $\beta$ -lactams, tetracyclines, aminoglycosides and macrolides. In addition, anti-TB drugs including first-line drugs (such as rifampicin and isoniazid) as well as some second-line agents (such as capreomycin) are ineffective against this pathogen (Table 1). Thus, the term "incurable nightmare" is often used to describe *M. abscessus* (Brown-Elliott et al., 2012; Nessar et al., 2012). So far, no reliable antibiotic regimen has been established for *M. abscessus* pulmonary disease. Antibiotic administration is largely empirical and relies on *in vitro* antibiotic susceptibility testing and definitive subspecies identification (Griffith et al., 2007; Ryu et al., 2016). Treatment of *M. abscessus* pulmonary disease as recommended by the British Thoracic Society involves administration of a multi-drug regimen encompassing intravenous antibiotics - amikacin, tigecycline, and imipenem along with an oral macrolide (clarithromycin or azithromycin) for clinical isolates susceptible to macrolides, during the initial treatment phase. For the continuation phase of treatment, nebulised amikacin and an oral macrolide combined with one to three of the following oral antibiotics: linezolid, clofazimine, minocycline, co-trimoxazole, and moxifloxacin are generally recommended (Haworth et al., 2017). Standard of care calls for continuation of antibiotic therapy for a minimum of 12 months after culture conversion. However, microbiological eradication of *M. abscessus* bacilli from lung tissues, using recommended antibiotic regimens, is rare and recurrence is often after completion of a treatment course and successful symptom management (Jarand et al., 2011; Stout et al., 2016).

As a result of extensive, repeated or inappropriate use of macrolides and aminoglycosides, which inhibit protein biosynthesis by binding to the large and small ribosomal subunits, respectively, *M. abscessus* strains with clinically acquired pan-macrolide and pan-aminoglycoside resistance have emerged, due to mutation(s) in the corresponding 23S (*rrl*) and 16S (*rrs*) rRNA genes (Wallace et al., 1996; Prammananan et al., 1998; Maurer et al., 2012; Nessar et al., 2012). Acquired resistance to macrolides and aminoglycosides severely limits the remaining treatment options and highlights the urgent need for new antimicrobial agents.

Mechanisms underpinning intrinsic drug resistance of *M. abscessus* are multi-fold and fall into two main groups: first, the presence of a highly impermeable cell envelope

**TABLE 1 |** Drug susceptibility of *Mycobacterium abscessus* wild-type and isogenic mutants in comparison to screening concentrations for *M. tuberculosis* with decreased susceptibility.

Antimicrobial agent	<i>M. tuberculosis</i> * (mg/L)	<i>M. abscessus</i> MIC (mg/L)	<i>M. abscessus</i> mutant MIC (mg/L)	Reference
Isoniazid	0.1	>512	<i>katG</i> <sup>+</sup> ( <i>M. tb</i> ): 32	Pers. Communication
Rifampicin	1.0	128	$\Delta$ <i>arr</i> : 0.25	Rominski et al., 2017a
Ethambutol	5.0	64 (polymorphism in target gene)	N.A.	Alcaide et al., 1997
Capreomycin	2.5	>256	$\Delta$ <i>eis2</i> : 4	Rominski et al., 2017c
Clarithromycin	N.D.	64	$\Delta$ <i>erm</i> : 0.5	Choi et al., 2012
Kanamycin B	N.D.	8	$\Delta$ <i>aac</i> (2'): 0.125	Rominski et al., 2017c
Amikacin	1.0	4	$\Delta$ <i>eis2</i> : 0.25	Rominski et al., 2017c
Streptomycin	1.0	32	$\Delta$ <i>str</i> (3''): 2	Dal Molin et al., 2017
Tetracycline	N.D.	60	$\Delta$ <i>tetX</i> : 4	Rudra et al., 2018
Amoxicillin	N.D.	>256	$\Delta$ <i>bla</i> : 8	Dub��e et al., 2015
Ampicillin	N.D.	>256	$\Delta$ <i>bla</i> : 4	Dub��e et al., 2015

Epidemiologic cutoff values (ECOFF; MGIT960) as defined by Cambau et al. (2015); MIC, minimal inhibitory concentration; N.A., Not available; N.D., Not determined.

and/or multi-drug efflux pumps might reduce the effective concentration of antibiotics within the bacterial cells; second, the genome of *M. abscessus* encodes several putative enzymes which can inactivate antibiotics by modification and/or degradation or lower the affinity of the drug for its target by modifying the target (Ripoll et al., 2009; Nessar et al., 2012). For long, molecular investigations aimed at elucidating antibiotic resistance mechanisms of *M. abscessus* were limited, however, significant progress has been made in recent years owing to the development of efficient tools for genetic manipulation of this bacterium (Figure 1). In this article, we provide an up-to-date overview of the main molecular mechanisms of antibiotic resistance in *M. abscessus* with a focus on the drug-target-modifying or drug-modifying enzymes and discuss the potential impact on clinical treatment. We also discuss how this knowledge could facilitate the discovery and development of new improved antimicrobial agents or help rescue the function of currently available antibiotics against *M. abscessus* and provide an update on the latest research underway to combat multi-drug resistant *M. abscessus* infections.

## ANTIBIOTIC-TARGET-MODIFYING ENZYMES IN *M. abscessus*

### Macrolide Resistance: MAB\_2297 [*erm*(41)] Encoded Erythromycin Ribosome Methylase

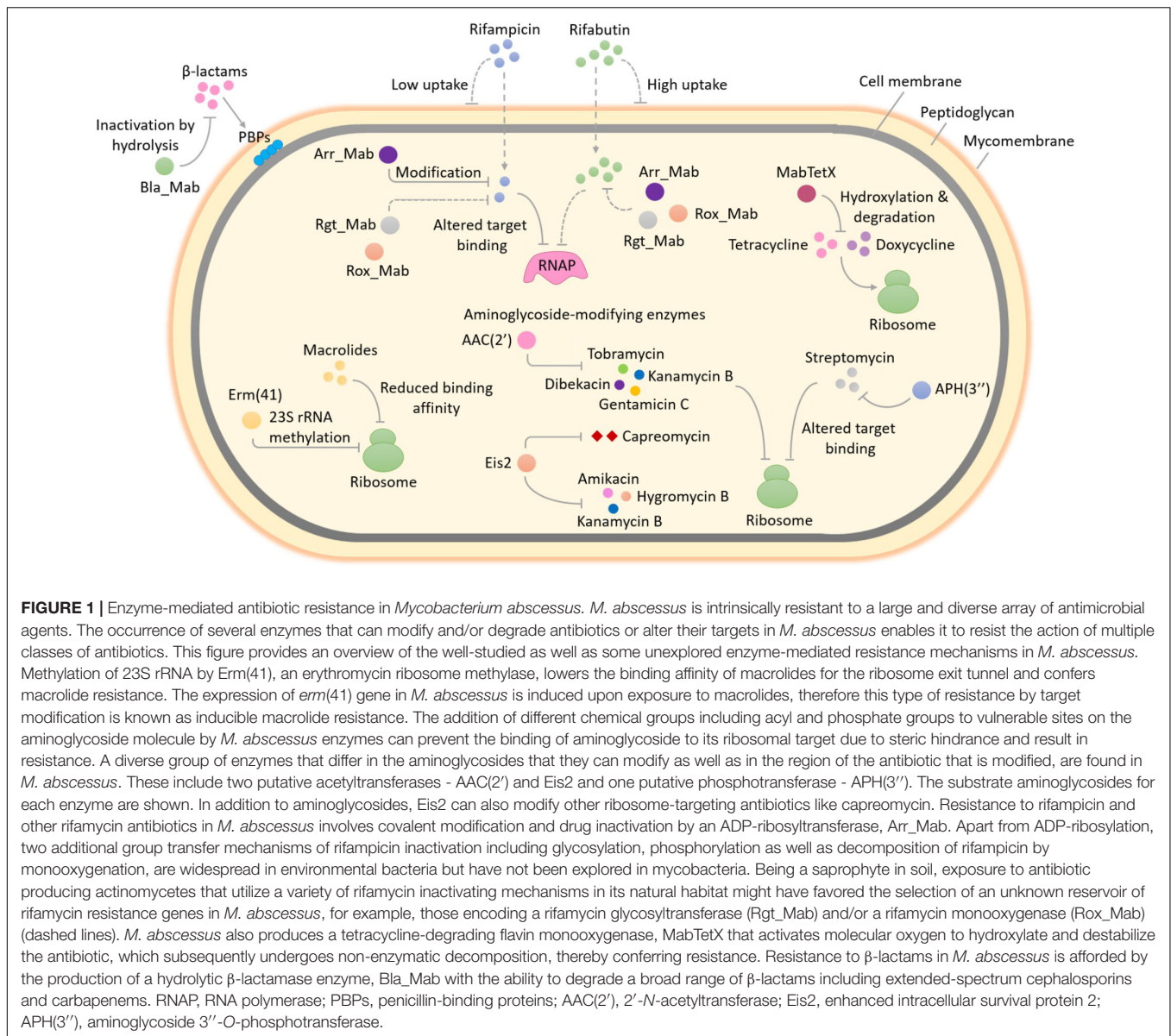
Macrolides are among the most successful antibiotics in the world and are highly prescribed for infections caused by non-tuberculous mycobacteria. A multidrug regimen recommended for *M. abscessus* infections almost always includes macrolides, particularly clarithromycin or azithromycin. Macrolides target the nascent polypeptide exit tunnel within the 50S ribosomal subunit, which accommodates the newly synthesized polypeptide chain as it emerges from the ribosome. Following macrolide binding, the elongation of polypeptide chain continues up to a few amino acids before the peptidyl-tRNA dissociates

from the ribosome thereby arresting protein synthesis (Wilson, 2014).

In bacteria, innate resistance towards macrolides is most often the result of increased efflux, ribosomal methyltransferases and GTP-dependent macrolide kinases (Pawlowski et al., 2018). A well-studied example of macrolide resistance by target modification in bacteria is the erythromycin ribosome methylase (Erm) enzyme, which specifically methylates A2058 nucleotide of 23S rRNA and lowers the macrolide affinity for ribosome exit tunnel (Blair et al., 2015). An Erm methyltransferase is also responsible for inducible macrolide resistance in *M. abscessus* (Figure 1). Interestingly, the *erm*(41) gene, first described by Nash et al. (2009), is only functional in two out of three subspecies of the *M. abscessus* complex. The subspecies *M. massiliense* harbors deletions in its *erm*(41) gene copy which render the Erm(41) enzyme inactive and the bacterium susceptible to macrolides. This may explain why patients with *M. massiliense* infections have better treatment outcomes than patients with *M. abscessus* or *M. bolletii* infections. To further complicate matters, a T/C polymorphism at position 28 of the *erm*(41) sequence determines the appearance of inducible macrolide resistance in *M. bolletii* and *M. abscessus* subspecies. Only those isolates that harbor a T28 *erm*(41) sequevar develop inducible macrolide resistance. Phenotypic detection of inducible macrolide resistance in the *M. abscessus* complex by drug susceptibility testing requires extended incubation (14 days rather than 3 days) in the presence of the drug (Nash et al., 2009; Kim et al., 2010).

A functional role of *erm*(41) in inducible resistance to macrolides has been confirmed by genetic studies. Deletion of *erm*(41) in *M. abscessus* lowered the minimal inhibitory concentration (MIC) of clarithromycin by 128-fold (Table 1), while introduction of a functional *M. abscessus* *erm*(41) gene into *M. massiliense* led to a 64-fold increase in the clarithromycin MIC on day 14 (Choi et al., 2012). Inducible Erm(41)-dependent rRNA methylation severely affects the efficacy of macrolides against *M. abscessus*. The long duration of partially effective macrolide therapy, eventually favors the emergence of isolates with high-level constitutive macrolide resistance. Mutations that alter the





23S rRNA nucleotides 2058 and 2059 within the peptidyl-transferase center (PTC), are a frequent cause of constitutive macrolide resistance in *M. abscessus* (Wallace et al., 1996; Bastian et al., 2011; Brown-Elliott et al., 2015).

## ANTIBIOTIC-MODIFYING ENZYMES IN *M. abscessus*

### Aminoglycoside Resistance: MAB\_4395 [*aac(2')*] & MAB\_4532c (*eis2*) Encoded Acetyltransferases; MAB\_2385 Encoded 3''-O-Phosphotransferase

Aminoglycosides are among the broadest classes of antibiotics that interfere with specific steps in bacterial protein synthesis

(Kohanski et al., 2010). All aminoglycosides primarily target the 16S rRNA component of 30S ribosomal subunit, however, their exact binding site and mode of action may differ slightly depending on their chemical structure. For example, 2-deoxystreptomine (2-DOS) aminoglycosides (such as kanamycin, amikacin, apramycin, arbekacin), which are chemically distinct from the non-DOS aminoglycosides (for example, streptomycin), specifically bind to the A-site region of small ribosomal subunit at the decoding center, encompassing conserved nucleotides A1492 and A1493 of 16S rRNA. 2-DOS aminoglycosides function by inhibiting the translocation of tRNA-mRNA complex through the ribosome. Additionally, several members of this class also induce protein mistranslation by promoting and stabilizing the interaction of non-cognate aminoacylated tRNAs with mRNA. In contrast, the non-DOS aminoglycoside streptomycin targets a different region of 16S rRNA in proximity of the decoding center

and mainly interferes with the delivery of aminoacylated tRNA to the A-site (Wilson, 2014).

Bacterial resistance to aminoglycosides may occur due to low cell permeability/efflux, mutations or modification of 16S rRNA, mutations in ribosomal protein genes and enzymatic drug modification. Drug and target modification being the most extensively studied and clinically relevant aminoglycoside resistance mechanisms in pathogenic bacteria. Covalent modification of key amino or hydroxyl groups within the aminoglycoside molecule by bacterial enzymes markedly decreases the binding affinity for the ribosomal target and confers aminoglycoside resistance. Aminoglycoside-modifying enzymes in bacteria can be grouped into three main classes namely, acetyltransferases, nucleotidyltransferases, and phosphotransferases (D'Costa et al., 2011; Blair et al., 2015). A worrying discovery that the genome of emerging pathogen *M. abscessus* encodes several putative aminoglycoside-modifying enzymes which include members of all three classes, came in 2009 when Ripoll et al first sequenced the complete genome of *M. abscessus* (Ripoll et al., 2009). Some predictions were later substantiated by drug susceptibility testing and biochemical analysis (Maurer et al., 2015). More recently, direct genetic evidence for a functional role of some of these putative aminoglycoside-modifying enzymes in *M. abscessus* intrinsic resistance to this antibiotic class, was obtained.

Similar to all other mycobacteria that have been studied so far (Aínsa et al., 1997), *M. abscessus* also harbors a 2'-N-acetyltransferase [AAC(2')], encoded by MAB\_4395 gene. This enzyme is capable of acetylating several aminoglycosides bearing a 2' amino group including gentamicin C, dibekacin, tobramycin and kanamycin B (Figure 1). This was evident upon deletion of MAB\_4395 in *M. abscessus* ATCC 19977 strain which led to a 4-64-fold reduction in the MICs of these aminoglycosides (Table 1) (Rominski et al., 2017c). Interestingly, another N-acetyltransferase capable of conferring aminoglycoside resistance in *M. abscessus* is a homolog of *Anabaena variabilis* enhanced intracellular survival (Eis) protein (Figure 1). Eis proteins are widespread in mycobacteria as well as other Gram-positive bacteria. Eis protein in *Mycobacterium tuberculosis* has been shown to enhance intracellular survival within macrophages by acetylating and activating dual-specificity protein phosphatase 16/mitogen-activated protein kinase phosphatase-7 (DUSP16/MPK-7), a JNK-specific phosphatase that inhibits inflammation, autophagy and subsequent death of infected macrophages (Shin et al., 2010; Kim et al., 2012). Furthermore, upregulation of *eis* gene expression owing to mutations in its promoter was associated with kanamycin resistance in one-third of clinical isolates encompassing a large collection of *M. tuberculosis* strains from diverse geographical locations (Zaunbrecher et al., 2009). More recently, structural and biochemical characterization of *M. tuberculosis* Eis revealed its unprecedented ability to acetylate multiple amines of several aminoglycosides (Chen et al., 2011).

Analysis of *M. abscessus* genome revealed two genes namely, MAB\_4124 (*eis1*) and MAB\_4532c (*eis2*), which show homology to *eis* from *M. tuberculosis* (Rv2416c) and *A. variabilis* (Ava\_4977), respectively. Drug susceptibility testing could

confirm a functional role for Eis2 but not Eis1, in intrinsic aminoglycoside resistance. Of note, deletion of MAB\_4532c enhanced *M. abscessus* susceptibility towards a heterogeneous group of aminoglycosides including the cornerstone drug amikacin as well as a non-aminoglycoside antibiotic capreomycin (Table 1) (Rominski et al., 2017c). Furthermore, heterologous expression of *eis2* in *Mycobacterium smegmatis* decreased amikacin susceptibility (Hurst-Hess et al., 2017). Capreomycin is a cyclic peptide antibiotic that inhibits protein synthesis by blocking peptidyl-tRNA translocation. The drug is particularly active against *M. tuberculosis* and commonly used as a second-line agent for the treatment of TB (Stanley et al., 2010; Wilson, 2014). Although remarkable, the discovery that *M. abscessus* Eis2 exhibits a broad-spectrum acetyltransferase activity is not a real surprise. In fact, recent structural and functional studies on Eis enzymes from *M. tuberculosis* (Eis\_Mtb), *M. smegmatis* (Eis\_Msm), and *A. variabilis* (Eis\_Ava) revealed the presence of an unusually large and complex active site unlike in other aminoglycoside acetyltransferases. The unique structure equips Eis with a regio-versatile multi-acetylating acetyltransferase activity towards a broad range of substrates (Chen et al., 2012; Pricer et al., 2012; Jennings et al., 2013). However, there seems to be a notable difference between the acetylation activities of *M. abscessus* Eis2 and *M. tuberculosis* Eis especially towards capreomycin as the MIC of capreomycin in *M. abscessus* is about 100-fold higher than that in *M. tuberculosis* and this difference is almost completely abolished by the deletion of *eis2* gene (Table 1) (Rominski et al., 2017c).

Aminoglycoside acetyltransferases Eis2 and AAC(2') do not appear to modify streptomycin, an aminoglycoside which exhibits moderate *in vitro* activity against *M. abscessus* (MIC: 32 mg/L) (Rominski et al., 2017c). Instead, a MAB\_2385 encoded putative aminoglycoside 3''-O-phosphotransferase is the main determinant for intrinsic streptomycin resistance in *M. abscessus* (Figure 1). Twelve putative aminoglycoside phosphotransferases are encoded in the genome of *M. abscessus* (Ripoll et al., 2009). Among them, MAB\_2385 was found to be a close homolog of *aph(3'')-Ic*, an aminoglycoside 3''-O-phosphotransferase gene which was previously shown to confer streptomycin resistance in *Mycobacterium fortuitum* (Ramón-García et al., 2006). Deletion of MAB\_2385 enhanced the susceptibility of *M. abscessus* towards streptomycin by 16-fold (Table 1). Furthermore, MAB\_2385 was able to confer streptomycin resistance when heterologously expressed in *M. smegmatis* (Dal Molin et al., 2017). To the best of our knowledge other annotated aminoglycoside phosphotransferases (such as MAB\_0163c, MAB\_0313c, MAB\_0327, MAB\_0951, and MAB\_1020) and aminoglycoside acetyltransferases (for example, MAB\_0247c, MAB\_0404c, MAB\_0745, MAB\_4235c, and MAB\_4324c) have not been addressed experimentally, however, interesting regulatory features of drug resistance genes have been elucidated.

A WhiB7 like protein encoded by MAB\_3508c was recently uncovered as a multi-drug inducible transcriptional regulator that modulates the expression of genes conferring aminoglycoside and macrolide resistance in *M. abscessus*. WhiB7, an autoregulatory transcriptional activator, is a determinant of

innate antibiotic resistance in mycobacteria. In *M. tuberculosis*, genes conferring antibiotic resistance including *ermMT* (Rv1988), *eis* (Rv2416c) and *tap* (Rv1258c) are induced in a *whiB7*-dependent manner, upon exposure to antibiotics. Similarly, the *erm*(41) and *eis2* genes in *M. abscessus* are included in the *whiB7* regulon (Hurst-Hess et al., 2017). A recent study by Pryjma et al. (2017) demonstrated that exposure to sub-inhibitory levels of clarithromycin, enhanced amikacin as well as clarithromycin resistance in *M. abscessus* in a *whiB7*-dependent manner. Of note, deletion of MAB\_3508c rendered *M. abscessus* more susceptible to amikacin (fourfold) and clarithromycin (eightfold), while complementation with an intact gene copy restored resistance to these antibiotics, suggesting a role of *whiB7* in *M. abscessus* amikacin and clarithromycin resistance. Interestingly, pre-exposure to clarithromycin, which is a potent inducer of *M. abscessus whiB7*, enhanced the resistance of *M. abscessus* wild-type strain towards amikacin by fourfold but had no effect on amikacin susceptibility of a  $\Delta whiB7$  mutant. In addition, pre-treatment with clarithromycin markedly increased resistance to itself in the wild-type strain but did not alter clarithromycin sensitivity of a  $\Delta whiB7$  mutant. Furthermore, a quantitative reverse transcription-PCR assay revealed a *whiB7*-dependent upregulation of *eis2* and *erm*(41) genes following pre-exposure to clarithromycin. These observations support the following conclusions: (i) in *M. abscessus*, *whiB7* (MAB\_3508c) activates the expression of genes which confer amikacin and clarithromycin resistance, i.e., *eis2* and *erm*(41), (ii) strong induction of *whiB7* following exposure to clarithromycin confers cross-resistance to amikacin in addition to activating inducible macrolide resistance (Pryjma et al., 2017). Thus, *in vitro* findings by Pryjma et al. (2017) suggest that front-line antibiotics, amikacin and clarithromycin may exhibit antagonistic effects when used in combination for the treatment of *M. abscessus* infections. This may at least partially explain the limited clinical efficacy of currently recommended multi-drug regimen for *M. abscessus* which almost always includes these two antibiotics.

## Tetracycline Resistance: MAB\_1496c Encoded Flavin Monooxygenase

Tetracyclines are a class of broad-spectrum natural product antibiotics that interfere with bacterial protein synthesis. These drugs bind with high affinity to the small ribosomal subunit and specifically interfere with the delivery of aminoacylated tRNA to the A-site (Kohanski et al., 2010; Wilson, 2014). While tetracyclines are an important class of ribosome-targeting antibiotics, their anthropogenic and prolific use in the clinic and agriculture has led to emergence of resistance among benign as well as pathogenic bacteria, thereby limiting their efficacy. Despite growing resistance, tetracyclines continue to remain one of the most successful and widely used chemotherapeutics against bacterial infections (Allen et al., 2010). Furthermore, a renaissance for tetracyclines is being fuelled by the development of next-generation derivatives such as tigecycline, approved for clinical use in 2005 and omadacycline and eravacycline, undergoing late-phase clinical trials (Kinch and Patridge, 2014). In fact, tigecycline is the only new antibiotic and the sole

member of its class, which has been introduced in the treatment of recalcitrant *M. abscessus* lung disease with clinical evidence (Wallace et al., 2014).

Previously, resistance towards tetracyclines was thought to be exclusively mediated by two mechanisms namely, drug efflux and ribosome protection. However, evidence for enzymatic inactivation of tetracyclines by a family of flavin adenine dinucleotide (FAD)-dependent monooxygenases in both benign and pathogenic bacteria has been recently documented. Few well characterized examples of such tetracycline-modifying enzymes include TetX from the obligate anaerobe *Bacteroides fragilis* and Tet(56) found in human pathogen *Legionella longbeachae*, the causative agent of Legionnaires' disease and Pontiac fever (Yang et al., 2004; Park et al., 2017). Whether such a mechanism of tetracycline resistance existed in mycobacteria was not known until recently, when Rudra and colleagues demonstrated through elegant experiments, that high levels of intrinsic resistance towards tetracycline and second generation derivative doxycycline in *M. abscessus* was mediated by a flavin monooxygenase, MabTetX (Figure 1). Deletion of MAB\_1496c (encoding a putative FAD binding monooxygenase) enhanced the susceptibility of *M. abscessus* towards tetracycline and doxycycline by 15–20-fold (Table 1), while a complemented  $\Delta$ MAB\_1496c strain exhibited even higher levels of resistance towards both antibiotics compared to the wild-type strain, supporting the notion that MAB\_1496c is a primary determinant for *M. abscessus* intrinsic tetracycline resistance. Furthermore, UV-Visible and mass spectrometry assays with purified MAB\_1496c protein provided evidence for hydroxylation and subsequent degradation of tetracycline and doxycycline by MabTetX (Rudra et al., 2018). Interestingly, MAB\_1496c is not a member of the *whiB7* regulon, even though tetracycline is a strong inducer of *whiB7* in *M. abscessus* (Hurst-Hess et al., 2017). In fact, MabTetX expression is regulated by a TetR family repressor, MabTetR<sub>x</sub> (encoded by the upstream gene MAB\_1497c) which represses the MAB\_1497c-MAB\_1496c operon by binding to a 35bp operator sequence *tetO*. This repression is relieved in the presence of tetracycline and doxycycline which can bind the repressor protein thereby preventing its interaction with the *tetO* operator (Rudra et al., 2018).

The glycylicycline antibiotic tigecycline, specifically designed to evade resistance by ribosome protection and efflux, is not invulnerable to modification by *Bacteroides* TetX, which was recently also identified in numerous clinically relevant pathogens (Volkers et al., 2011; Leski et al., 2013). Although tigecycline was found to be a poor substrate of TetX, alarmingly enzymatic activity of TetX was substantially improved upon acquisition of single amino acid substitutions to confer resistance at clinically relevant tigecycline concentrations when overexpressed in *Escherichia coli* (Linkevicius et al., 2016). Surprisingly, tigecycline resisted MabTetX-dependent degradation and did not induce its expression, which might contribute to its excellent *in vitro* activity and moderate clinical efficacy against *M. abscessus* (Rudra et al., 2018). However, selective pressure exerted by tigecycline will eventually favor



the emergence of *M. abscessus* isolates carrying extended-spectrum FAD-monooxygenases with the ability to modify glycolylcyclines.

Another interesting feature is the ability of anhydrotetracycline (ATc), a degradation product of tetracycline with poor antibiotic activity which was recently shown to be a competitive inhibitor of TetX and Tet(56) flavin monooxygenases, to rescue antibiotic activities of tetracycline and doxycycline against *M. abscessus* expressing MabTetX (Park et al., 2017; Rudra et al., 2018). However, ATc was also found to induce MabTetX expression by binding the MabTetR<sub>x</sub> repressor, a property at odds with its MabTetX inhibition activity. While this finding together with the severe side effects associated with ATc use, makes it an unattractive drug candidate, it nevertheless represents a flexible starting point for developing MabTetX inhibitors with improved activity, better tolerability and minimal binding affinity towards MabTetR<sub>x</sub>. Co-administration with a potent MabTetX monooxygenase inhibitor can potentially rescue the clinical efficacies of tetracycline and doxycycline which are currently inactive against the *M. abscessus* complex (Burgos et al., 2011; Rudra et al., 2018).

## β-Lactam Resistance: MAB\_2875 Encoded β-Lactamase

β-lactams constitute an important class of antibiotics that function by inhibiting cell wall synthesis in bacteria. The β-lactam structure closely resembles the terminal D-alanyl-D-alanine dipeptide of peptidoglycan, a substrate of penicillin binding proteins (also known as D, D-transpeptidases) that catalyze the final cross-linking step of peptidoglycan synthesis. Penicilloylation of the D, D-transpeptidase active site following treatment with β-lactams, disables the enzyme and prevents the formation of peptidoglycan cross-links, thereby interfering with cell wall biosynthesis (Kohanski et al., 2010). Despite being the cornerstones of antimicrobial chemotherapy and constituting one of the largest groups of antibiotics available today, only two members of the β-lactam class namely, cefoxitin (a cephalosporin) and imipenem (a carbapenem) form a part of the antibiotic arsenal for *M. abscessus*. Furthermore, cefoxitin and imipenem have been shown to exhibit only moderate *in vitro* activity against *M. abscessus* with MICs of 32 and 4 mg/L, respectively (Lavollay et al., 2014). Majority of the other β-lactam antibiotics are not effective against this bacterium.

Several possible mechanisms may contribute to β-lactam resistance in *M. abscessus*. Low mycomembrane permeability and β-lactamase production may reduce the effective concentration of β-lactams at the site of action. Alternatively, modification of transpeptidase profile of the cell by replacement of penicillin binding proteins (D, D-transpeptidases) with L, D-transpeptidases for instance, may reduce the activity of penicillins and cephalosporins that have lower affinity for L, D-transpeptidase (Wivagg et al., 2014). However, most of the abovementioned resistance mechanisms are relatively unexplored in this organism except for one. Recently it was shown that the genome of *M. abscessus* encodes a strong, constitutive class A β-lactamase (Bla\_Mab) which renders it highly resistant towards most

β-lactams (Figure 1). The *M. abscessus* β-lactamase Bla\_Mab, encoded by MAB\_2875 is endowed with an exceptional broad-spectrum activity and can effectively hydrolyse several members of first- and second-generation cephalosporins, carbapenems, and penams (Soroka et al., 2014). The currently recommended β-lactams namely imipenem and cefoxitin are substrates of Bla\_Mab, however, they are hydrolysed at a very slow rate, which may contribute to their clinical efficacy. The instability of imipenem complicates drug susceptibility testing when generation times of test bacteria require incubations for several days as in the case of *M. abscessus* (Rominski et al., 2017b).

One strategy to overcome the hurdle of chromosomally encoded β-lactamase in *M. abscessus* is to co-administer a β-lactamase inhibitor with the failing β-lactam. Soroka and colleagues evaluated the *in vitro* efficacy of nitrocefin and meropenem in combination with approved β-lactamase inhibitors clavulanate, tazobactam, and sulbactam against Bla\_Mab. Surprisingly, Bla\_Mab inhibition was not detected for all the three inhibitor drugs (Soroka et al., 2014). This remarkable ability of Bla\_Mab to resist inhibition even surpasses that of BlaC, a class A β-lactamase encoded by its relative *M. tuberculosis*. Although BlaC is able to reverse inhibition caused by tazobactam and sulbactam, and return to its native functional form, clavulanate can slowly but irreversibly inhibit this enzyme (Sagar et al., 2017). In fact, a recent demonstration of *in vitro* synergy between meropenem and clavulanate in 13 extensively drug resistant (XDR) *M. tuberculosis* strains raised a renewed interest in β-lactams for use in TB therapy (Hugonnet et al., 2009). Fortunately, a surprising finding that *M. abscessus* Bla\_Mab is effectively disabled by the newly approved β-lactamase inhibitor avibactam provides a new hope for old antibiotics. A study by Dubée et al. (2015) showed that co-administering a small amount of avibactam (4 mg/L) with each of several representative antibiotics from three important β-lactam sub-classes (carbapenems, penams, and cephalosporins) lowered their MICs in *M. abscessus* CIP104536 strain to levels comparable to those observed in a β-lactamase deficient mutant. The authors also reported synergy between avibactam and amoxicillin within macrophages as well as in a zebrafish model of *M. abscessus* infection (Dubée et al., 2015). More recently, an *in vitro* synergy screen of 110 β-lactam - β-lactamase inhibitor combinations against an *M. abscessus* clinical isolate identified six potential hits. Five selected β-lactamase inhibitors encompassing both β-lactam-based (clavulanate, tazobactam, and sulbactam) as well as non-β-lactam-based inhibitors (avibactam and vaborbactam) were evaluated for synergistic effects with a large panel of β-lactams. Importantly, all observed synergy combinations exclusively involved non-β-lactam-based inhibitors, either avibactam or vaborbactam. Combinations of avibactam with each of the following drugs including ampicillin, amoxicillin, tebipenem, and panipenem displayed synergistic effects against *M. abscessus*. Furthermore, the carbapenems, tebipenem, and panipenem also exhibited synergy with a novel boronic acid β-lactamase inhibitor, vaborbactam (Aziz et al., 2018). Thus, non-β-lactam-based β-lactamase inhibitors such as avibactam and vaborbactam can potentially extend the spectrum

of  $\beta$ -lactams employed for the treatment of largely incurable, chronic *M. abscessus* pulmonary infections.

## Rifamycin Resistance: MAB\_0591 Encoded ADP-Ribosyltransferase

Rifampicin, a rifamycin antibiotic commonly employed as a first-line agent in the treatment of *M. tuberculosis* infections, hardly exhibits any activity towards the *M. abscessus* complex (MIC: 128 mg/L) (Table 1). Since rifamycins inhibit transcription by binding with high affinity to *rpoB* encoded  $\beta$ -subunit of bacterial RNA polymerase, resistance towards this group of antibiotics is most commonly conferred by point mutations within the target gene *rpoB*, especially in *M. tuberculosis* (Campbell et al., 2001). However, additional rifamycin resistance mechanisms that involve enzymatic drug inactivation are also prevalent in other mycobacterial species. The presence of a rifamycin ADP-ribosyltransferase in *M. smegmatis* was known for quite a while (Quan et al., 1997), however, only recently genetic studies involving gene knockout and heterologous expression unveiled the existence of a MAB\_0591 encoded ADP-ribosyltransferase as the major determinant for high levels of innate rifamycin resistance in *M. abscessus* (Figure 1). Deletion of MAB\_0591 enhanced the susceptibility of *M. abscessus* towards three tested rifamycins including rifaximin, rifapentine and rifampicin by 64–512-fold, respectively (Rominski et al., 2017a).

Surprisingly, a rifampicin derivative, rifabutin showed up as an attractive hit (MIC: 2.5 mg/L), when a set of 2,700 FDA-approved drugs were screened against a clinical isolate of *M. abscessus*. Rifabutin exhibited approximately 10-fold higher potency *in vitro* relative to rifampicin and rifapentine, against reference strains representative of all three subspecies of the *M. abscessus* complex as well as a collection of clinical isolates and its bactericidal activity was comparable to or better than that of clarithromycin, one of the cornerstone drugs for *M. abscessus* infections (Aziz et al., 2017). The higher efficacy of rifabutin relative to rifampicin in *M. abscessus* as well as in *M. tuberculosis* (with no known rifamycin inactivating enzymes) suggests its increased accumulation within the bacterial cells. This may be due to differences in membrane penetration or specificity of efflux systems towards different rifamycins (Figure 1). Based on the findings reported by Aziz et al. (2017), rifampicin analog rifabutin, appears to be a potential candidate for repurposing and its clinical efficacy in patients with *M. abscessus* pulmonary disease should be further explored.

## CLINICAL RELEVANCE

Although the inherent ability of *M. abscessus* to exhibit resistance to a wide array of antibiotics has long been recognized, our knowledge of the prodigious diversity of mechanisms involved has improved immensely in recent years, owing to the development of tools for genetic manipulation of *M. abscessus*. A better understanding of the genetic basis of innate antibiotic resistance is a prerequisite for the discovery and development of synergistic drug combinations, where one agent serves to salvage the antibacterial activity of a failing antibiotic by inhibiting an

intrinsic resistance mechanism. This is exemplified by the recent discovery that avibactam is a potent inhibitor of *M. abscessus*  $\beta$ -lactamase Bla\_Mab. When used in combination, avibactam lowered the MICs of several  $\beta$ -lactams in *M. abscessus* by 4–32-fold (Dub  e et al., 2015; Kaushik et al., 2017). Despite the success of  $\beta$ -lactamase inhibitors, the concept of designing drugs which target intrinsic resistance mechanisms in bacteria, has not gained substantial clinical exploitation beyond this antibiotic class (Brown, 2015). Recently, a repertoire of drug-modifying and target-modifying enzymes conferring innate resistance to aminoglycosides, tetracyclines, rifamycins, and macrolides in *M. abscessus* were delineated (Nash et al., 2009; Dal Molin et al., 2017; Rominski et al., 2017a,c; Rudra et al., 2018). Likewise, attempts to identify chemical entities which could potentially rescue the function of one or more key members from each aforementioned antibiotic group, should be pursued in the near future.

Since the discovery that mutations within *eis* promoter are associated with kanamycin resistance in *M. tuberculosis*, vigorous attempts to identify Eis inhibitors that can potentially rescue kanamycin activity in this pathogen have been pursued with notable success. Recently, several structurally diverse competitive Eis inhibitors with 1,2,4-triazino[5,6*b*]indole-3-thioether-based, sulfonamide-based, pyrrolo[1,5-*a*]pyrazine-based and isothiazole S,S-dioxide heterocyclic scaffolds were identified by high-throughput screenings of large compound libraries. Importantly, few promising inhibitors could completely restore kanamycin susceptibility in a kanamycin resistant *M. tuberculosis* strain *in vitro* at concentrations that were non-cytotoxic to mammalian cells (Green et al., 2012; Garzan et al., 2016, 2017; Willby et al., 2016; Ngo et al., 2018). Remarkably, inhibitors designed for *M. tuberculosis* Eis, were also found to be active against Eis homologs in *M. smegmatis* and *A. variabilis* (Chen et al., 2012; Pricer et al., 2012). These findings give rise to an intriguing possibility that Eis\_Mtb inhibitors may be able to overcome Eis2 mediated resistance towards aminoglycoside and/or non-aminoglycoside antibiotics like capreomycin and restore their effectiveness against *M. abscessus*. Hence efforts to explore the activity of Eis\_Mtb inhibitors against *M. abscessus* Eis2 enzyme should be undertaken. Alternatively, extending drug discovery efforts towards targeting WhiB7, could be a possible strategy to tackle intrinsic macrolide and aminoglycoside resistance in *M. abscessus*. WhiB7 is a conserved transcriptional regulator in mycobacteria that co-ordinates intrinsic resistance to a wide range of ribosome-targeting drugs (Burian et al., 2012). Recent evidence suggests that genes which contribute to intrinsic aminoglycoside and macrolide resistance in *M. abscessus*, i.e., *eis2* and *erm*(41), are induced in a *whiB7*-dependent manner upon exposure to ribosomal antibiotics (Hurst-Hess et al., 2017). Thus, with the aim to kill two birds with one stone, compounds that prevent the induction of *eis2* and *erm*(41) resistance genes in *M. abscessus* by modulating the synthesis and/or activity of WhiB7, should be sought.

Since the discovery and development of compounds that can circumvent existing resistance mechanisms in *M. abscessus* is a daunting and time-consuming task, an immediate and relatively simple solution to improve treatment efficacy may

involve repurposing and repositioning of currently available antibiotics. A number of candidate drugs suited for this purpose have been reviewed in detail elsewhere (Wu et al., 2018) and are therefore only briefly mentioned here. For example, a systematic screening of a collection of FDA-approved drugs revealed rifabutin, an analog of rifampicin, exhibiting potent *in vitro* activity against *M. abscessus* (MIC: 2.5 mg/L) (Aziz et al., 2017). Similarly, clofazimine, a leprosy drug currently repurposed for treatment of TB, was also found to be active against *M. abscessus*. In addition to promising *in vitro* activity (MIC  $\leq$  1 mg/L), clofazimine displayed adequate clinical efficacy and safety when evaluated in patients with *M. abscessus* pulmonary infections (Yang et al., 2017). Likewise, a novel oxazolidinone LCB01-0371, presently in Phase II clinical development for TB, exhibited high potency against *M. abscessus* compared to linezolid, an FDA-approved drug of the same class (Kim et al., 2017). Moreover, some aminoglycosides such as kanamycin A, apramycin, isepamicin, and arbekacin were also found to exhibit potent *in vitro* activity against *M. abscessus* (MICs  $\leq$  1 mg/L) (Rominski et al., 2017c). Besides these, other compounds that have been extensively studied for their inhibitory effects against *M. abscessus* encompass anti-TB drug bedaquiline, some TB actives like indole-2-carboxamides and piperidinol-based compound 1 (PIPD1), as well as second generation thiacetazone derivatives like D6, D15, and D17 (Dupont et al., 2016, 2017; Franz et al., 2017; Halloum et al., 2017; Kozikowski et al., 2017).

In addition, several recent studies have reported *in vitro* synergies between novel combinations of existing antibiotic classes. For example, vancomycin, an inhibitor of cell wall synthesis, exhibited synergy with clarithromycin, an inhibitor of protein synthesis, in *M. abscessus* (Mukherjee et al., 2017). Likewise, a combination of tigecycline (which inhibits protein synthesis) with teicoplanin (which disrupts peptidoglycan synthesis) displayed a strong synergistic effect against *M. abscessus* (Aziz et al., 2018). Since the mycobacterial cell envelope forms a major permeability barrier, drugs that perturb cell wall integrity, are likely to synergize with drugs having intracellular targets (Le Run et al., 2018). Such combinations based on mechanistic drug action should be further explored for synergistic effects against *M. abscessus*. For example, PIPD1 and indole-2-carboxamides, which disrupt mycolic acid synthesis by targeting MmpL3 transporter in mycobacteria (Dupont et al., 2016; Franz et al., 2017), may be able to potentiate the effects of other drugs modulating intracellular targets.

Taken together, a huge reservoir of intrinsic resistance genes combined with an unusually high evolutionary potential of its genome to acquire resistance renders almost all available antibiotics ineffective against *M. abscessus*. Therefore, in the long run, a better understanding of natural resistance mechanisms in *M. abscessus* will help jump start drug discovery projects for (i) compounds that can rescue current antibiotics by neutralizing intrinsic resistance (termed antibiotic resistance breakers), (ii) new drugs with one or more improved properties like ability to resist enzymatic modification and/or degradation, high affinity for modified or mutated targets, better solubility/uptake and low propensity for efflux (Brown, 2015). In the short run,

available knowledge from studies seeking synergy between antibiotics together with those investigating potential candidates for repurposing, will aid in the development of dosing regimens that display adequate clinical efficacy and resistance to which will only emerge slowly.

## OUTLOOK AND PERSPECTIVES

Molecular genetic tools that allow investigators to systematically probe gene function in mycobacteria by generation of unmarked gene deletions and functional complementation, have proven extremely useful in dissecting the intrinsic resistome of *M. abscessus* while genetic investigations in other NTM species are still lagging behind. Using the available genetic 'toolkit,' 7 orthologs of known resistance gene families have been discovered so far in *M. abscessus* that provide resistance or 'immunity' against antibiotics targeting major cellular pathways including cell wall synthesis ( $\beta$ -lactams), RNA synthesis (rifamycins) and protein synthesis (macrolides, aminoglycosides, and tetracyclines). The impressive resistance diversity shown by *M. abscessus* is expected given its natural habitat is shared by many antibiotic producers, primarily the soil dwelling actinomycetes which are also reservoirs of extensively diverse resistance elements (Pawlowski et al., 2016; Crofts et al., 2017; Koteva et al., 2018). Exposure to a variety of noxious antimicrobial molecules in the soil might have favored selection of specialized and diverse resistance mechanisms in *M. abscessus* as well as in other NTM species.

Till now, the approach that researchers used to systematically probe the intrinsic resistome of *M. abscessus* involved analysis of few selected genes identified based on homology to known resistance determinants from other bacteria. As a result, this approach mainly revealed genes within *M. abscessus* that were involved in previously known resistance mechanisms and disguised the presence of potential determinants of entirely new resistance mechanisms that lack characterized homologs in other bacteria. To capture the remarkable resistance diversity of *M. abscessus* as well as other environmental NTM with unprecedented depth, future studies in this field should incorporate next-generation approaches that provide a comprehensive genome-wide definition of loci required for antibiotic resistance, using cutting-edge technologies. One such technique which can efficiently mine resistance determinants with novel sequences as well as assign resistance functions to known genes that were not previously shown to be involved in drug resistance, is the most recent incarnation of insertional mutagenesis, called transposon insertion sequencing (TIS) (Chao et al., 2016).

The utility of TIS in elucidating novel antibiotic resistance functions is highlighted by its recent application to the discovery of a repertoire of previously unknown intrinsic factors impacting resistance to antibiotics of different classes including oxazolidinones, fluoroquinolones, and aminoglycosides in pathogens of high clinical concern such as *Staphylococcus aureus*, *P. aeruginosa* and *E. coli* (Gallagher et al., 2011; Shan et al., 2015; Rajagopal et al., 2016). Screening large collections



of transposon insertion mutants for antibiotic susceptibility using TIS approach, has identified promising novel targets for drugs that can restore or enhance susceptibility to existing antibiotics. For example, analysis of insertion mutants spanning all non-essential *S. aureus* genes for fitness defects resulting from exposure to antibiotics, identified pathways/genes including *graRS* and *vraFG* (*graRS/vraFG*), *fmtA*, *mprF*, *SAOUHSC\_01025*, and *SAOUHSC\_01050* which if inhibited, can greatly improve the efficacy of existing antibiotics including daptomycin, vancomycin, gentamicin, ciprofloxacin, oxacillin, and linezolid and extend their utility for treating *S. aureus* infections (Rajagopal et al., 2016).

In the coming years, high-throughput next-generation strategies like TIS will undoubtedly rise to prominence in the NTM drug resistance field and radically advance our understanding of resistance mechanisms in *M. abscessus* and other emerging NTM pathogens. Such breakthrough technologies will greatly impact the kinds of questions which can be addressed in this field, for example, which resistance elements unique to *M. abscessus* render it unusually more drug resistant than other NTM species? or does *M. abscessus* already

carry genes which confer resistance to novel NTM drugs in development?

## AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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## REFERENCES

- Aínsa, J. A., Pérez, E., Pelicic, V., Berthet, F. X., Gicquel, B., and Martín, C. (1997). Aminoglycoside 2'-N-acetyltransferase genes are universally present in mycobacteria: characterization of the *aac*(2')-Ic gene from *Mycobacterium tuberculosis* and the *aac*(2')-Id gene from *Mycobacterium smegmatis*. *Mol. Microbiol.* 24, 431–441.
- Alcaide, F., Pfyffer, G. E., and Telenti, A. (1997). Role of *embB* in natural and acquired resistance to ethambutol in Mycobacteria. *Antimicrob. Agents Chemother.* 41, 2270–2273.
- Allen, H. K., Donato, J., Wang, H. H., Cloud-Hansen, K. A., Davies, J., and Handelsman, J. (2010). Call of the wild: antibiotic resistance genes in natural environments. *Nat. Rev. Microbiol.* 8, 251–259. doi: 10.1038/nrmicro.2312
- Aziz, D. B., Low, J. L., Wu, M.-L., Gengenbacher, M., Teo, J. W. P., Dartois, V., et al. (2017). Rifabutin is active against *Mycobacterium abscessus* complex. *Antimicrob. Agents Chemother.* 61:e00155–17. doi: 10.1128/AAC.00155-17
- Aziz, D. B., Teo, J. W. P., Dartois, V., and Dick, T. (2018). Teicoplanin - tigecycline combination shows synergy against *Mycobacterium abscessus*. *Front. Microbiol.* 9:932. doi: 10.3389/fmicb.2018.00932
- Bastian, S., Veziris, N., Roux, A.-L., Brossier, F., Gaillard, J.-L., Jarlier, V., et al. (2011). Assessment of clarithromycin susceptibility in strains belonging to the *Mycobacterium abscessus* group by *erm*(41) and *rrl* sequencing. *Antimicrob. Agents Chemother.* 55, 775–781. doi: 10.1128/AAC.00861-10
- Blair, J. M. A., Webber, M. A., Baylay, A. J., Ogbolu, D. O., and Piddock, L. J. V. (2015). Molecular mechanisms of antibiotic resistance. *Nat. Rev. Microbiol.* 13, 42–51. doi: 10.1038/nrmicro3380
- Brown, D. (2015). Antibiotic resistance breakers: can repurposed drugs fill the antibiotic discovery void? *Nat. Rev. Drug Discov.* 14, 821–832. doi: 10.1038/nrd4675
- Brown-Elliott, B. A., Nash, K. A., and Wallace, R. J. (2012). Antimicrobial susceptibility testing, drug resistance mechanisms, and therapy of infections with nontuberculous mycobacteria. *Clin. Microbiol. Rev.* 25, 545–582. doi: 10.1128/CMR.05030-11
- Brown-Elliott, B. A., Vasireddy, S., Vasireddy, R., Iakhiaeva, E., Howard, S. T., Nash, K., et al. (2015). Utility of sequencing the *erm*(41) gene in isolates of *Mycobacterium abscessus* subsp. *abscessus* with low and intermediate clarithromycin MICs. *J. Clin. Microbiol.* 53, 1211–1215. doi: 10.1128/JCM.02950-14
- Brown-Elliott, B. A., and Wallace, R. J. (2002). Clinical and taxonomic status of pathogenic nonpigmented or late-pigmenting rapidly growing Mycobacteria. *Clin. Microbiol. Rev.* 15, 716–746. doi: 10.1128/CMR.15.4.716-746.2002
- Bryant, J. M., Grogono, D. M., Rodriguez-Rincon, D., Everall, I., Brown, K. P., Moreno, P., et al. (2016). Population-level genomics identifies the emergence and global spread of a human transmissible multidrug-resistant nontuberculous mycobacterium. *Science* 354, 751–757. doi: 10.1126/science.aaf.8156
- Burgos, M. I., Fernández, R. A., Celej, M. S., Rossi, L. I., Fidelio, G. D., and Dassie, S. A. (2011). Binding of the highly toxic tetracycline derivative, anhydrotetracycline, to bovine serum albumin. *Biol. Pharm. Bull.* 34, 1301–1306. doi: 10.1248/bpb.34.1301
- Burian, J., Ramón-García, S., Howes, C. G., and Thompson, C. J. (2012). WhiB7, a transcriptional activator that coordinates physiology with intrinsic drug resistance in *Mycobacterium tuberculosis*. *Expert Rev. Anti Infect. Ther.* 10, 1037–1047. doi: 10.1586/eri.12.90
- Cambau, E., Viveiros, M., Machado, D., Raskine, L., Ritter, C., Tortoli, E., et al. (2015). Revisiting susceptibility testing in MDR-TB by a standardized quantitative phenotypic assessment in a European multicentre study. *J. Antimicrob. Chemother.* 70, 686–696. doi: 10.1093/jac/dku438
- Campbell, E. A., Korzheva, N., Mustaev, A., Murakami, K., Nair, S., Goldfarb, A., et al. (2001). Structural mechanism for rifampicin inhibition of bacterial RNA polymerase. *Cell* 104, 901–912. doi: 10.1016/S0092-8674(01)00286-0
- Chan, E., and Iseman, M. (2013). Underlying host risk factors for nontuberculous Mycobacterial lung disease. *Semin. Respir. Crit. Care Med.* 34, 110–123. doi: 10.1055/s-0033-1333573
- Chao, M. C., Abel, S., Davis, B. M., and Waldor, M. K. (2016). The design and analysis of transposon insertion sequencing experiments. *Nat. Rev. Microbiol.* 14, 119–128. doi: 10.1038/nrmicro.2015.7
- Chen, W., Biswas, T., Porter, V. R., Tsodikov, O. V., and Garneau-Tsodikova, S. (2011). Unusual regioversatility of acetyltransferase Eis, a cause of drug resistance in XDR-TB. *Proc. Natl. Acad. Sci.* 108, 9804–9808. doi: 10.1073/pnas.1105379108
- Chen, W., Green, K. D., Tsodikov, O. V., and Garneau-Tsodikova, S. (2012). The aminoglycoside multi-acetylating activity of the enhanced intracellular survival (Eis) protein from *Mycobacterium smegmatis* and its inhibition. *Biochemistry* 51, 4959–4967. doi: 10.1021/bi3004473
- Choi, G.-E., Shin, S. J., Won, C.-J., Min, K.-N., Oh, T., Hahn, M.-Y., et al. (2012). Macrolide treatment for *Mycobacterium abscessus* and *Mycobacterium massiliense* infection and inducible resistance. *Am. J. Respir. Crit. Care Med.* 186, 917–925. doi: 10.1164/rccm.201111-2005OC

- Choo, S. W., Wee, W. Y., Ngeow, Y. F., Mitchell, W., Tan, J. L., Wong, G. J., et al. (2014). Genomic reconnaissance of clinical isolates of emerging human pathogen *Mycobacterium abscessus* reveals high evolutionary potential. *Sci. Rep.* 4:4061. doi: 10.1038/srep04061
- Crofts, T. S., Gasparrini, A. J., and Dantas, G. (2017). Next-generation approaches to understand and combat the antibiotic resistome. *Nat. Rev. Microbiol.* 15, 422–434. doi: 10.1038/nrmicro.2017.28
- Dal Molin, M., Gut, M., Rominski, A., Haldimann, K., Becker, K., and Sander, P. (2017). Molecular mechanisms of intrinsic streptomycin resistance in *Mycobacterium abscessus*. *Antimicrob. Agents Chemother.* 62:e01427-17. doi: 10.1128/AAC.01427-17
- D'Costa, V. M., King, C. E., Kalan, L., Morar, M., Sung, W. W. L., Schwarz, C., et al. (2011). Antibiotic resistance is ancient. *Nature* 477, 457–461. doi: 10.1038/nature10388
- Dubée, V., Bernut, A., Cortes, M., Lesne, T., Dorcène, D., Lefebvre, A.-L., et al. (2015).  $\beta$ -Lactamase inhibition by avibactam in *Mycobacterium abscessus*. *J. Antimicrob. Chemother.* 70, 1051–1058. doi: 10.1093/jac/dku510
- Dupont, C., Viljoen, A., Dubar, F., Blaise, M., Bernut, A., Pawlik, A., et al. (2016). A new piperidinol derivative targeting mycolic acid transport in *Mycobacterium abscessus*. *Mol. Microbiol.* 101, 515–529. doi: 10.1111/mmi.13406
- Dupont, C., Viljoen, A., Thomas, S., Roquet-Banères, F., Herrmann, J.-L., Pethe, K., et al. (2017). Bedaquiline inhibits the ATP synthase in *Mycobacterium abscessus* and is effective in infected zebrafish. *Antimicrob. Agents Chemother.* 61, 1–15. doi: 10.1128/AAC.01225-17
- Franz, N. D., Belardinelli, J. M., Kaminski, M. A., Dunn, L. C., Calado Nogueira de Moura, V., Blaha, M. A., et al. (2017). Design, synthesis and evaluation of indole-2-carboxamides with pan anti-mycobacterial activity. *Bioorg. Med. Chem.* 25, 3746–3755. doi: 10.1016/j.bmc.2017.05.015
- Gallagher, L. A., Shendure, J., and Manoil, C. (2011). Genome-scale identification of resistance functions in *Pseudomonas aeruginosa* using Tn-seq. *mBio* 2:e00315-10. doi: 10.1128/mBio.00315-10
- Garzan, A., Willby, M. J., Green, K. D., Gajadeera, C. S., Hou, C., Tsodikov, O. V., et al. (2016). Sulfonamide-based inhibitors of aminoglycoside acetyltransferase Eis abolish resistance to kanamycin in *Mycobacterium tuberculosis*. *J. Med. Chem.* 59, 10619–10628. doi: 10.1021/acs.jmedchem.6b01161
- Garzan, A., Willby, M. J., Ngo, H. X., Gajadeera, C. S., Green, K. D., Holbrook, S. Y. L., et al. (2017). Combating enhanced intracellular survival (Eis)-mediated kanamycin resistance of *Mycobacterium tuberculosis* by novel pyrrolo[1,5-a]pyrazine-based Eis inhibitors. *ACS Infect. Dis.* 3, 302–309. doi: 10.1021/acsinfectdis.6b00193
- Green, K. D., Chen, W., and Garneau-Tsodikova, S. (2012). Identification and characterization of inhibitors of the aminoglycoside resistance acetyltransferase Eis from *Mycobacterium tuberculosis*. *ChemMedChem* 7, 73–77. doi: 10.1002/cmdc.201100332
- Griffith, D. E., Aksamit, T., Brown-Elliott, B. A., Catanzaro, A., Daley, C., Gordin, F., et al. (2007). An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. *Am. J. Respir. Crit. Care Med.* 175, 367–416. doi: 10.1164/rccm.200604-571ST
- Halloum, I., Viljoen, A., Khanna, V., Craig, D., Bouchier, C., Brosch, R., et al. (2017). Resistance to thiacetazone derivatives active against *Mycobacterium abscessus* involves mutations in the MmpL5 transcriptional repressor MAB\_4384. *Antimicrob. Agents Chemother.* 61, e02509-16. doi: 10.1128/AAC.02509-16
- Haworth, C. S., Banks, J., Capstick, T., Fisher, A. J., Gorsuch, T., Laurenson, I. F., et al. (2017). British thoracic society guideline for the management of non-tuberculous mycobacterial pulmonary disease (NTM-PD). *BMJ Open Respir. Res.* 4:e000242. doi: 10.1136/bmjresp-2017-000242
- Hoefsloot, W., van Ingen, J., Andrejak, C., Ängeby, K., Bauriaud, R., Bemer, P., et al. (2013). The geographic diversity of nontuberculous mycobacteria isolated from pulmonary samples: an NTM-NET collaborative study. *Eur. Respir. J.* 42, 1604–1613. doi: 10.1183/09031936.00149212
- Howard, S. T. (2006). Spontaneous reversion of *Mycobacterium abscessus* from a smooth to a rough morphotype is associated with reduced expression of glycopeptidolipid and reacquisition of an invasive phenotype. *Microbiology* 152, 1581–1590. doi: 10.1099/mic.0.28625-0
- Hugonnet, J., Tremblay, L. W., Boshoff, H. I., Barry, C. E., and Blanchard, J. S. (2009). Meropenem-clavulanate is effective against extensively drug-resistant *Mycobacterium tuberculosis*. *Science* 323, 1215–1218. doi: 10.1126/science.1167498
- Hurst-Hess, K., Rudra, P., and Ghosh, P. (2017). *Mycobacterium abscessus* WhiB7 regulates a species-specific repertoire of genes to confer extreme antibiotic resistance. *Antimicrob. Agents Chemother.* 61, e01347-17. doi: 10.1128/AAC.01347-17
- Jarand, J., Levin, A., Zhang, L., Huitt, G., Mitchell, J. D., and Daley, C. L. (2011). Clinical and microbiologic outcomes in patients receiving treatment for *Mycobacterium abscessus* pulmonary disease. *Clin. Infect. Dis.* 52, 565–571. doi: 10.1093/cid/ciq237
- Jennings, B. C., Labby, K. J., Green, K. D., and Garneau-Tsodikova, S. (2013). Redesign of substrate specificity and identification of aminoglycoside binding residues of Eis from *Mycobacterium tuberculosis*. *Biochemistry* 52, 5125–5132. doi: 10.1021/bi4002985
- Kaushik, A., Gupta, C., Fisher, S., Story-Roller, E., Galanis, C., Parrish, N., et al. (2017). Combinations of avibactam and carbapenems exhibit enhanced potencies against drug-resistant *Mycobacterium abscessus*. *Future Microbiol.* 12, 473–480. doi: 10.2217/fmb-2016-0234
- Kim, H.-Y., Kim, B. J., Kook, Y., Yun, Y.-J., Shin, J. H., Kim, B.-J., et al. (2010). *Mycobacterium massiliense* is differentiated from *Mycobacterium abscessus* and *Mycobacterium bolletii* by erythromycin ribosome methyltransferase gene (erm) and clarithromycin susceptibility patterns. *Microbiol. Immunol.* 54, 347–353. doi: 10.1111/j.1348-0421.2010.00221.x
- Kim, K. H., An, D. R., Song, J., Yoon, J. Y., Kim, H. S., Yoon, H. J., et al. (2012). *Mycobacterium tuberculosis* Eis protein initiates suppression of host immune responses by acetylation of DUSP16/MKP-7. *Proc. Natl. Acad. Sci. U.S.A.* 109, 7729–7734. doi: 10.1073/pnas.1120251109
- Kim, T. S., Choe, J. H., Kim, Y. J., Yang, C., Kwon, H., Jeong, J., et al. (2017). Activity of LCB01-0371, a novel oxazolidinone, against *Mycobacterium abscessus*. *Antimicrob. Agents Chemother.* 61, e02752-16. doi: 10.1128/AAC.02752-16
- Kinch, M. S., and Patridge, E. (2014). An analysis of FDA-approved drugs for infectious disease: antibacterial agents. *Drug Discov. Today* 19, 1510–1513. doi: 10.1016/j.drudis.2014.05.012
- Kohanski, M. A., Dwyer, D. J., and Collins, J. J. (2010). How antibiotics kill bacteria: from targets to networks. *Nat. Rev. Microbiol.* 8, 423–435. doi: 10.1038/nrmicro2333
- Koteva, K., Cox, G., Kelso, J. K., Surette, M. D., Zubyk, H. L., Ejim, L., et al. (2018). Rox, a rifamycin resistance enzyme with an unprecedented mechanism of action. *Cell Chem. Biol.* 25, 403–412.e5. doi: 10.1016/j.chembiol.2018.01.009
- Kozikowski, A. P., Onajole, O. K., Stec, J., Dupont, C., Viljoen, A., Richard, M., et al. (2017). Targeting mycolic acid transport by indole-2-carboxamides for the treatment of *Mycobacterium abscessus* infections. *J. Med. Chem.* 60, 5876–5888. doi: 10.1021/acs.jmedchem.7b00582
- Lavollay, M., Dubée, V., Heym, B., Herrmann, J.-L., Gaillard, J.-L., Gutmann, L., et al. (2014). In vitro activity of cefoxitin and imipenem against *Mycobacterium abscessus* complex. *Clin. Microbiol. Infect.* 20, O297–O300. doi: 10.1111/1469-0691.12405
- Le Run, E., Arthur, M., and Mainardi, J.-L. (2018). In vitro and intracellular activity of imipenem combined to rifabutin and avibactam against *Mycobacterium abscessus*. *Antimicrob. Agents Chemother.* 62:e00623-18. doi: 10.1128/AAC.00623-18
- Lee, M.-R., Sheng, W.-H., Hung, C.-C., Yu, C.-J., Lee, L.-N., and Hsueh, P.-R. (2015). *Mycobacterium abscessus* complex infections in humans. *Emerg. Infect. Dis.* 21, 1638–1646. doi: 10.3201/2109.141634
- Leski, T. A., Bangura, U., Jimmy, D. H., Ansumana, R., Lizewski, S. E., Stenger, D. A., et al. (2013). Multidrug-resistant tet(X)-containing hospital isolates in Sierra Leone. *Int. J. Antimicrob. Agents* 42, 83–86. doi: 10.1016/j.ijantimicag.2013.04.014
- Linkevicius, M., Sandegren, L., and Andersson, D. I. (2016). Potential of tetracycline resistance proteins to evolve tigecycline resistance. *Antimicrob. Agents Chemother.* 60, 789–796. doi: 10.1128/AAC.02465-15



- Maurer, F. P., Bruderer, V. L., Castelberg, C., Ritter, C., Scherbakov, D., Bloemberg, G. V., et al. (2015). Aminoglycoside-modifying enzymes determine the innate susceptibility to aminoglycoside antibiotics in rapidly growing mycobacteria. *J. Antimicrob. Chemother.* 70, 1412–1419. doi: 10.1093/jac/dku550
- Maurer, F. P., Ruegger, V., Ritter, C., Bloemberg, G. V., and Böttger, E. C. (2012). Acquisition of clarithromycin resistance mutations in the 23S rRNA gene of *Mycobacterium abscessus* in the presence of inducible erm(41). *J. Antimicrob. Chemother.* 67, 2606–2611. doi: 10.1093/jac/dks279
- Mukherjee, D., Wu, M.-L., Teo, J. W. P., and Dick, T. (2017). Vancomycin and clarithromycin show synergy against *Mycobacterium abscessus* in vitro. *Antimicrob. Agents Chemother.* 61, e01298-17. doi: 10.1128/AAC.01298-17
- Nash, K. A., Brown-Elliott, A. B., and Wallace, R. J. (2009). A novel gene, erm(41), confers inducible macrolide resistance to clinical isolates of *Mycobacterium abscessus* but is absent from *Mycobacterium chelonae*. *Antimicrob. Agents Chemother.* 53, 1367–1376. doi: 10.1128/AAC.01275-08
- Nessar, R., Cambau, E., Reyat, J. M., Murray, A., and Gicquel, B. (2012). *Mycobacterium abscessus*: a new antibiotic nightmare. *J. Antimicrob. Chemother.* 67, 810–818. doi: 10.1093/jac/dkr578
- Ngo, H. X., Green, K. D., Gajadeera, C. S., Willby, M. J., Holbrook, S. Y. L., Hou, C., et al. (2018). Potent 1,2,4-triazino[5,6b]indole-3-thioether inhibitors of the kanamycin resistance enzyme Eis from *Mycobacterium tuberculosis*. *ACS Infect. Dis.* 4, 1030–1040. doi: 10.1021/acscinfdis.8b00074
- Park, J., Gasparrini, A. J., Reck, M. R., Symister, C. T., Elliott, J. L., Vogel, J. P., et al. (2017). Plasticity, dynamics, and inhibition of emerging tetracycline-resistance enzymes. *Nat. Chem. Biol.* 13, 730–736. doi: 10.1038/nchembio.2376
- Pawlowski, A. C., Stogios, P. J., Koteva, K., Skarina, T., Evdokimova, E., Savchenko, A., et al. (2018). The evolution of substrate discrimination in macrolide antibiotic resistance enzymes. *Nat. Commun.* 9:112. doi: 10.1038/s41467-017-02680-0
- Pawlowski, A. C., Wang, W., Koteva, K., Barton, H. A., McArthur, A. G., and Wright, G. D. (2016). A diverse intrinsic antibiotic resistome from a cave bacterium. *Nat. Commun.* 7:13803. doi: 10.1038/ncomms13803
- Prammananan, T., Sander, P., Brown, B. A., Frischkorn, K., Onyi, G. O., Zhang, Y., et al. (1998). A single 16S ribosomal RNA substitution is responsible for resistance to amikacin and other 2-deoxystreptamine aminoglycosides in *Mycobacterium abscessus* and *Mycobacterium chelonae*. *J. Infect. Dis.* 177, 1573–1581. doi: 10.1086/515328
- Pricer, R. E., Houghton, J. L., Green, K. D., Mayhoub, A. S., and Garneau-Tsodikova, S. (2012). Biochemical and structural analysis of aminoglycoside acetyltransferase Eis from *Anabaena variabilis*. *Mol. Biosyst.* 8, 3305–3313. doi: 10.1039/c2mb25341k
- Pryjma, M., Burian, J., Kuchinski, K., and Thompson, C. J. (2017). Antagonism between front-line antibiotics clarithromycin and amikacin in the treatment of *Mycobacterium abscessus* infections is mediated by the whiB7 gene. *Antimicrob. Agents Chemother.* 61, e01353-17. doi: 10.1128/AAC.01353-17
- Quan, S., Venter, H., and Dabbs, E. R. (1997). Ribosylative inactivation of rifampin by *Mycobacterium smegmatis* is a principal contributor to its low susceptibility to this antibiotic. *Antimicrob. Agents Chemother.* 41, 2456–2460.
- Rajagopal, M., Martin, M. J., Santiago, M., Lee, W., Kos, V. N., Meredith, T., et al. (2016). Multidrug intrinsic resistance factors in *Staphylococcus aureus* identified by profiling fitness within high-diversity transposon libraries. *mBio* 7: e00950-16. doi: 10.1128/mBio.00950-16
- Ramón-García, S., Otal, I., Martín, C., Gomez-Lus, R., and Ainsa, J. A. (2006). Novel streptomycin resistance gene from *Mycobacterium fortuitum*. *Antimicrob. Agents Chemother.* 50, 3920–3922. doi: 10.1128/AAC.00223-06
- Ripoll, F., Pasek, S., Schenowitz, C., Dossat, C., Barbe, V., Rottman, M., et al. (2009). Non mycobacterial virulence genes in the genome of the emerging pathogen *Mycobacterium abscessus*. *PLoS One* 4:e5660. doi: 10.1371/journal.pone.0005660
- Rominski, A., Roditscheff, A., Selchow, P., Böttger, E. C., and Sander, P. (2017a). Intrinsic rifamycin resistance of *Mycobacterium abscessus* is mediated by ADP-ribosyltransferase MAB\_0591. *J. Antimicrob. Chemother.* 72, 376–384. doi: 10.1093/jac/dkw466
- Rominski, A., Schulthess, B., Müller, D. M., Keller, P. M., and Sander, P. (2017b). Effect of  $\beta$ -lactamase production and  $\beta$ -lactam instability on MIC testing results for *Mycobacterium abscessus*. *J. Antimicrob. Chemother.* 72, 3070–3078. doi: 10.1093/jac/dkx284
- Rominski, A., Selchow, P., Becker, K., Brülle, J. K., Dal Molin, M., and Sander, P. (2017c). Elucidation of *Mycobacterium abscessus* aminoglycoside and capreomycin resistance by targeted deletion of three putative resistance genes. *J. Antimicrob. Chemother.* 72, 2191–2200. doi: 10.1093/jac/dkx125
- Rudra, P., Hurst-Hess, K., Lappierre, P., and Ghosh, P. (2018). High levels of intrinsic tetracycline resistance in *Mycobacterium abscessus* are conferred by a tetracycline-modifying monooxygenase. *Antimicrob. Agents Chemother.* 62:e00119-18. doi: 10.1128/AAC.00119-18
- Ryu, Y. J., Koh, W.-J., and Daley, C. L. (2016). Diagnosis and treatment of nontuberculous mycobacterial lung disease: clinicians' perspectives. *Tuberc. Respir. Dis.* 79, 74–84. doi: 10.4046/trd.2016.79.2.74
- Sagar, A., Haleem, N., Bashir, Y. M., and Ashish, A. (2017). Search for non-lactam inhibitors of mtb  $\beta$ -lactamase led to its open shape in apo state: new concept for antibiotic design. *Sci. Rep.* 7:6204. doi: 10.1038/s41598-017-06023-3
- Sanguinetti, M., Ardito, F., Fiscarelli, E., La Sorda, M., D'Argenio, P., Ricciotti, G., et al. (2001). Fatal pulmonary infection due to multidrug-resistant *Mycobacterium abscessus* in a patient with cystic fibrosis. *J. Clin. Microbiol.* 39, 816–819. doi: 10.1128/JCM.39.2.816-819.2001
- Shan, Y., Lazinski, D., Rowe, S., Camilli, A., and Lewis, K. (2015). Genetic basis of persister tolerance to aminoglycosides in *Escherichia coli*. *mBio* 6:e00078-15. doi: 10.1128/mBio.00078-15
- Shin, D.-M., Jeon, B.-Y., Lee, H.-M., Jin, H. S., Yuk, J.-M., Song, C.-H., et al. (2010). *Mycobacterium tuberculosis* Eis regulates autophagy, inflammation, and cell death through redox-dependent signaling. *PLoS Pathog.* 6:e1001230. doi: 10.1371/journal.ppat.1001230
- Soroka, D., Dubée, V., Soulier-Escrihueta, O., Cuinet, G., Hugonnet, J.-E., Gutmann, L., et al. (2014). Characterization of broad-spectrum *Mycobacterium abscessus* class A  $\beta$ -lactamase. *J. Antimicrob. Chemother.* 69, 691–696. doi: 10.1093/jac/dkt410
- Stanley, R. E., Blaha, G., Grodzicki, R. L., Strickler, M. D., and Steitz, T. A. (2010). The structures of the anti-tuberculosis antibiotics viomycin and capreomycin bound to the 70S ribosome. *Nat. Struct. Mol. Biol.* 17, 289–293. doi: 10.1038/nsmb.1755
- Stout, J. E., Koh, W.-J., and Yew, W. W. (2016). Update on pulmonary disease due to non-tuberculous mycobacteria. *Int. J. Infect. Dis.* 45, 123–134. doi: 10.1016/j.ijid.2016.03.006
- Tortoli, E., Kohl, T. A., Brown-Elliott, B. A., Trovato, A., Leão, S. C., Garcia, M. J., et al. (2016). Emended description of *Mycobacterium abscessus*, *Mycobacterium abscessus* subsp. *abscessus* and *Mycobacterium abscessus* subsp. *bolletii* and designation of *Mycobacterium abscessus* subsp. *massiliense* comb. nov. *Int. J. Syst. Evol. Microbiol.* 66, 4471–4479. doi: 10.1099/ijsem.0.001376
- Volkers, G., Palm, G. J., Weiss, M. S., Wright, G. D., and Hinrichs, W. (2011). Structural basis for a new tetracycline resistance mechanism relying on the TetX monooxygenase. *FEBS Lett.* 585, 1061–1066. doi: 10.1016/j.febslet.2011.03.012
- Wallace, R. J., Dukart, G., Brown-Elliott, B. A., Griffith, D. E., Scerpella, E. G., and Marshall, B. (2014). Clinical experience in 52 patients with tigecycline-containing regimens for salvage treatment of *Mycobacterium abscessus* and *Mycobacterium chelonae* infections. *J. Antimicrob. Chemother.* 69, 1945–1953. doi: 10.1093/jac/dku062
- Wallace, R. J., Meier, A., Brown, B. A., Zhang, Y., Sander, P., Onyi, G. O., et al. (1996). Genetic basis for clarithromycin resistance among isolates of *Mycobacterium chelonae* and *Mycobacterium abscessus*. *Antimicrob. Agents Chemother.* 40, 1676–1681.

- Willby, M. J., Green, K. D., Gajadeera, C. S., Hou, C., Tsodikov, O. V., Posey, J. E., et al. (2016). Potent inhibitors of acetyltransferase Eis Overcome kanamycin resistance in *Mycobacterium tuberculosis*. *ACS Chem. Biol.* 11, 1639–1646. doi: 10.1021/acscchembio.6b00110
- Wilson, D. N. (2014). Ribosome-targeting antibiotics and mechanisms of bacterial resistance. *Nat. Rev. Microbiol.* 12, 35–48. doi: 10.1038/nrmicro3155
- Wivagg, C. N., Bhattacharyya, R. P., and Hung, D. T. (2014). Mechanisms of  $\beta$ -lactam killing and resistance in the context of *Mycobacterium tuberculosis*. *J. Antibiot.* 67, 645–654. doi: 10.1038/ja.2014.94
- Wu, M.-L., Aziz, D. B., Dartois, V., and Dick, T. (2018). NTM drug discovery: status, gaps and the way forward. *Drug Discov. Today* 23, 1502–1519. doi: 10.1016/j.drudis.2018.04.001
- Yang, B., Jhun, B. W., Moon, S. M., Lee, H., Park, H. Y., Jeon, K., et al. (2017). Clofazimine-containing regimen for the treatment of *Mycobacterium abscessus* lung disease. *Antimicrob. Agents Chemother.* 61:e02052-16. doi: 10.1128/AAC.02052-16
- Yang, W., Moore, I. F., Koteva, K. P., Bareich, D. C., Hughes, D. W., and Wright, G. D. (2004). TetX Is a Flavin-dependent monooxygenase conferring resistance to tetracycline antibiotics. *J. Biol. Chem.* 279, 52346–52352. doi: 10.1074/jbc.M409573200
- Zaunbrecher, M. A., Sikes, R. D., Metchock, B., Shinnick, T. M., and Posey, J. E. (2009). Overexpression of the chromosomally encoded aminoglycoside acetyltransferase Eis confers kanamycin resistance in *Mycobacterium tuberculosis*. *Proc. Natl. Acad. Sci. U.S.A.* 106, 20004–20009. doi: 10.1073/pnas.0907925106

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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